

Synthesis of a Novel C₂₆ Marine Sterol

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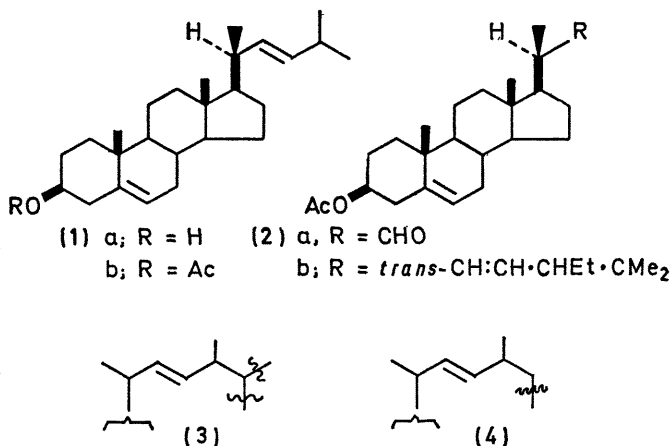
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Summary The synthesis of 22-*trans*-26,27-bisnor-ergosta-5,22-dien-3 β -ol by an unequivocal route establishes this as the sterol isolated from several Pelecypoda by Idler and his co-workers.

reductive work-up (Zn-HOAc) and treatment with saturated NaHSO₃ solution gave 81% of the bisulphite

RECENTLY Idler and his co-workers suggested structure (1a) for the C₂₆ sterol isolated from the scallop, *Placopecten magellanicus* (Gmelin). The sterol also occurs in several other Pelecypoda including the blue mussel, *Mytilus edulis* L.; the clam, *Mya arenaria* L.; the ocean quahog, *Arctica islandica* L.; and the oyster, *Crassostera virginica* (Gmelin).¹ Because of the biogenetic novelty of the side-chain structure suggested for (1a) and the uncertainties associated with the stereochemistry at C-20 we have synthesized this sterol by an unequivocal route.

The synthesis of (1a) was accomplished *via* a Wittig reaction of the 20*R*-aldehyde (2a)² which was prepared from stigmasteryl acetate, (2b). Bromination of (2b) with iodobenzene dibromide at -5° in hexane gave 91% of 5 α ,6 β -dibromo-stigmast-22-en-3 β -yl acetate.† Ozonolysis of the bromosterol (-70° in CH₂Cl₂-pyridine) followed by



derivative of (2a). The bisulphite derivative was converted into (2a) (95%) by reaction with 10% Na₂CO₃.

† M.p.s are uncorrected. All new compounds had correct analyses and, where not specifically discussed, the expected spectroscopic data.

Reaction of (2a) with isobutyl-triphenylphosphorane (HI salt + BuLi) in diethyl ether (r t for 2 h then 60° for 12 h) followed by treatment with acetic anhydride in pyridine gave 40% of (1b) and its Δ^{22} -*cis*-isomer (1.4). Wittig reaction of (2b) in hexane⁴ reversed the Δ^{22} -*trans-cis*-ratio to 6:1. Several recrystallizations (MeOH) of the crude (1b) from the latter reaction were required to give pure 20S³ (1b), m p 142.5—143°. The n m r of (1b) gave singlets at δ 0.687 (18-H₃), 1.01 (19-H₃), 2.00 (Ac), and doublets at δ 0.925 (24-dimethyl) and 0.992 (21-H₃). Hydrolysis of (1b) in refluxing base (2% KOH in 10% H₂O-MeOH) gave (1a), m p 143—144°, $[\alpha]_D^{25}$ -65° (c, 2.7).

The mass spectrum of synthetic (1a) was identical with that published¹ for the C₂₆ sterol of Pelecypoda. The n m r spectrum of (1a) exhibited singlets at δ 0.70 (18-H₃) and 1.01 (19-H₃), doublets were observed at δ 1.01 (21-H₃)

and 0.96 (24-dimethyl), within experimental error of those reported by Idler¹.

We are most grateful to Dr. D. R. Idler for direct comparison of synthetic (1a) (m p 142—143°) with the natural sterol (m p 138—140°) by mixed m p (138—141°), i r, n m r, and g l p c ‡ all of which indicated the synthetic and natural C₂₆ sterols were identical.

The interesting side-chain of (1) could conceivably arise by degradation of a C₂₄ methylated sterol (3) or more interestingly, from degradation of a sterol (4) produced upon cyclization of a modified squalene. In the latter case one terminal isoprene unit must be attached in a head-to-head fashion.

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‡ Dr. Idler performed the g l p c analysis of the free C₂₆ sterols on a 6 ft 1% OV-1 column at 217°. Comparison of synthetic (1b) and the natural C₂₆ sterol acetate by g l p c was performed on a 12 ft 3% XE 60 and a 12 ft 3% NGS column at 210°.

¹ D. R. Idler, P. M. Wiseman, and L. M. Safe, *Steroids*, 1970, **16**, 451. Dr. Idler has informed us that the m p of the C₂₆ sterol should be reported as 138—140°.

² M. Fryberg, A. C. Oehlschlager, and A. M. Unrau, *Tetrahedron*, 1971, **27**, 1261.

³ D. H. R. Barton, T. Shouri, and D. A. Widdowson, *Chem Comm*, 1970, 940.

⁴ R. F. N. Hutchins, M. J. Thompson, and J. A. Svoboda, *Steroids*, 1969, **15**, 113.